

SHORT COMMUNICATIONS

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Purification and some properties of cytochrome *c* derived from the marine worm, *Dendrostomum zosteri*colum

We have isolated cytochrome *c* in a state of high purity from the marine worm, *Dendrostomum zosteri*colum, and studied some of its properties.

The marine worms (1500 g) were cut into pieces of approx. 1 cm length, homogenized with 0.01 M phosphate buffer, pH 7.0, and the resultant suspension was allowed to stand overnight at 5°. It was then filtered through celite and the filtrate centrifuged at $35000 \times g$ for 20 min. The supernatant fluid obtained was dialyzed against 0.01 M phosphate buffer, pH 7.0, and the dialysate was charged on an Amberlite CG-50 column which had been equilibrated with the same buffer as used for the dialysis. The cytochrome *c* was adsorbed on the column. After the column had been washed with 0.05 M phosphate buffer, pH 7.0, the cytochrome *c* was eluted with 0.1 M phosphate buffer, pH 7.0, containing 10 % saturated $(\text{NH}_4)_2\text{SO}_4$. To the eluate obtained (70 ml, $A_{550 \text{ m}\mu}$ (reduced) = 0.130) we added $(\text{NH}_4)_2\text{SO}_4$ to the saturation. The solution was centrifuged at $35000 \times g$ for 10 min. The supernatant fluid

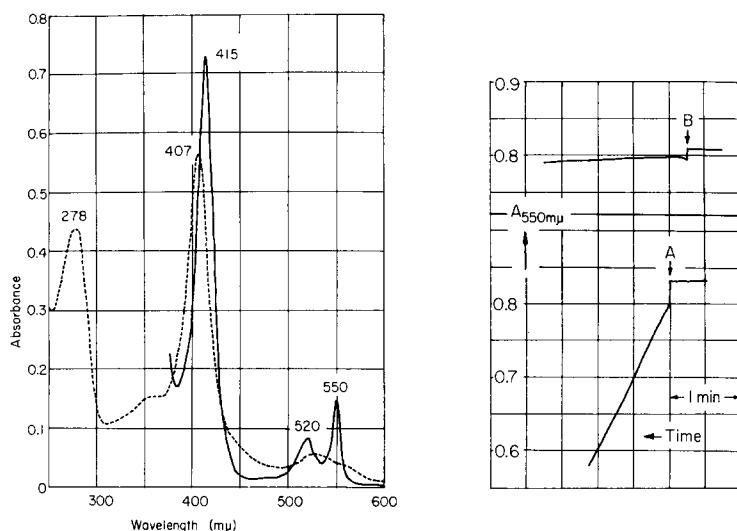


Fig. 1. Absorption spectrum of *D. zosteri*colum cytochrome *c*. The protein was dissolved in 0.2 M phosphate buffer, pH 7.0. ----, oxidized; —, reduced with $\text{Na}_2\text{S}_2\text{O}_4$.

Fig. 2. Reactivities of *D. zosteri*colum cytochrome *c* with *Pseudomonas* and cow cytochrome oxidases. The cytochrome *c* was reduced by addition of a small amount of $\text{Na}_2\text{S}_2\text{O}_4$ and dialyzed for several hours against 0.04 M phosphate buffer, pH 6.5. The reactions were carried out in 0.04 M phosphate buffer at pH 6.5 and at 21°. At points B and A, 0.05 ml of 5.6 μM *Pseudomonas* cytochrome oxidase and 0.05 ml of 5.2 μM cow cytochrome oxidase were added, respectively, to 1.0 ml of the cytochrome *c* solution.

was charged on an Amberlite CG-50 column (described above) after dialysis against 0.01 M phosphate buffer, pH 7.0. The cytochrome *c* was adsorbed on the column. After the column had been washed with 0.05 M phosphate buffer, pH 7.0, the cytochrome was eluted with 0.05 M phosphate buffer (pH 7.0) containing 20 % saturated $(\text{NH}_4)_2\text{SO}_4$. The eluate obtained was dialyzed against 0.04 M phosphate buffer, pH 6.5, overnight, and the resulting dialysate was used as the purified *D. zosteri* cytochrome *c* preparation. From 15 g of worms about 3 mg of cytochrome *c* was obtained, assuming that the ϵ_{mM} at 550 m μ and the mol. wt. were 28000 and 12000, respectively.

The cytochrome *c* possessed absorption maxima at 407 and 525 m μ in the oxidized form, at 415, 520 and 550 m μ in the reduced form, and its pyridine hemochrome showed absorption maxima at 414, 518 and 549 m μ .

The cytochrome *c* reacted very poorly with *Pseudomonas* cytochrome oxidase¹, and very rapidly with cow cytochrome oxidase². Thus, the molecular activities (moles of cytochrome *c* oxidized per mole of cytochrome oxidase per min) were 1.0 and 105 in the reactions with *Pseudomonas* and cow cytochrome oxidase, respectively, at 21° and pH 6.5. These molecular activities were relative values; that is, in the reaction with *Pseudomonas* cytochrome oxidase, the molecular activity obtained when *Pseudomonas aeruginosa* cytochrome *c* reacted with the bacterial enzyme was taken as 100 %, and in the reaction with cow cytochrome oxidase the molecular activity when *Saccharomyces ovoidormis* cytochrome *c* reacted with the animal enzyme was taken as 100 % (ref. 3). From the facts given above, it is apparent that *D. zosteri* cytochrome *c* is a mammalian-type cytochrome *c*.

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1 T. YAMANAKA, *Ann. Rep. Scient. Works, Fac. Sci., Osaka Univ.*, 11 (1963) 77.

2 K. OKUNUKI, in E. H. STOTZ AND M. FLORKIN, *Comprehensive Biochemistry*, Vol. 14, Elsevier, Amsterdam, 1966, p. 232.

3 T. YAMANAKA AND K. OKUNUKI, *J. Biol. Chem.*, 239 (1964) 1813.

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